Objective: To obtain knowledge and understanding about the production of antibiotics by making use of a variety of tests with bacteria from a soil isolate.

Research Question: Does our soil contain an undiscovered antibiotic producer that can fight against the increasing issue of drug-resistant infections?

Method: Serial Dilute 10g of soil with 90mL of water and repeat until there are plates with countable colonies. We used the 10-4 dilution plate for our countable colony [see figure 1]. We then picked and patched each of our bacteria to a new agar plate and incubated the bacteria for growth. After the growth of our master plate, we then picked and patched each bacterium to a new TSA media each with a different ESKAPE safe relative. After incubating, we screened the plates for antibiotic production with our single colonies. Of the sixteen different types of bacteria, we had two antibiotic producers [see figure 2] and a red bacteria that we chose to further analyze.

Procedures with Unknown #3:

PCR Indole Test Citrate Test Catalase Test

Triple Sugar Iron Test Oxidase Test Mitility agar Test Gel Electrophoresis

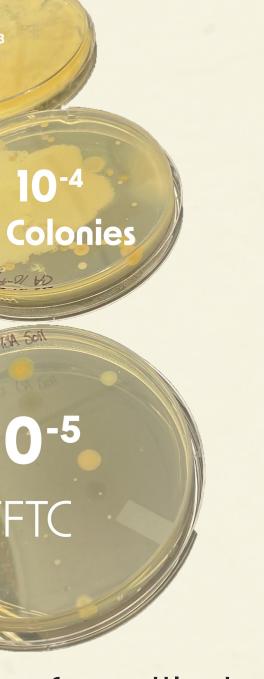
Master plate

Urease Reaction Gram Stain + KOH Endospore Stain

FIGURE 1

Unknown #3





TSA plates after dilution

Antibiotic Results



Only unknown #1 and #5 were found to produce antibiotics when exposed to *E. raffinosus*, leaving faint zones of inhibition.

Key Results for Unknown #3

BLAST Analysis: Serratia [inhibens] E Value = 0.0

Gram Stain: gram negative

Endospore Stain: negative

KOH Test: positive

Biochemical Testing



Negative: no

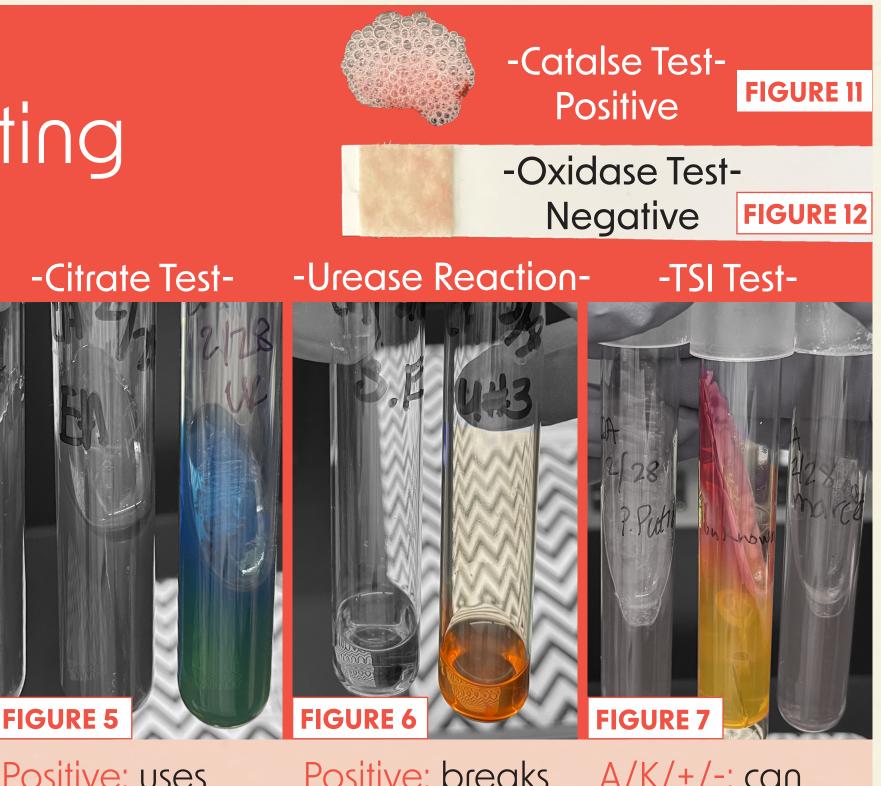
signs of indole

to breakdown

tryptophan



Positive: is a motile bacteria due to flagella



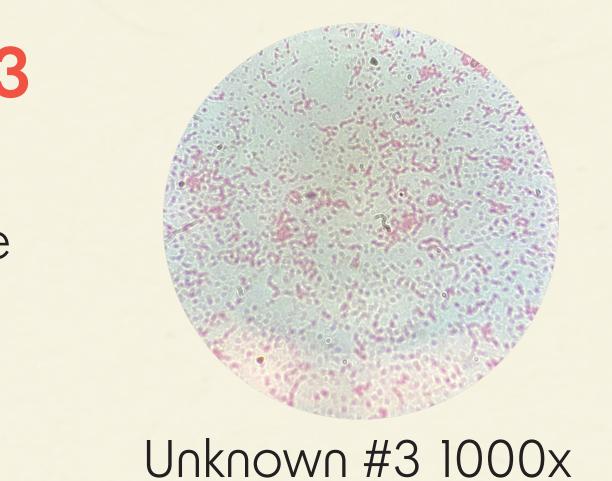
Positive: uses citrate as a source of carbon

Differential and Selective Media Testing

[<image/> <text></text>	<image/> <text></text>	<image/> <text></text>
	Growth +	Growth +	Growth +
	Colony Color:	Colony Color:	Colony Color:
	red/purple	metallic green	neon pink
	Medium Color:	Medium Color:	Medium Color:
	no change	no change	no change
	Conclusion:	Conclusion:	Conclusion:
	lactose fermenter	lactose fermenter	non-sugar ferm.

(the conclusively inconclusive bacteria)

FIGURE 2 A. baylyi E. raffinosus S. epidermidis P. putida



Positive: breaks down urea into ammonia with enzyme urease

4/K/+/-: can ferment glucose while making hydrogen gas



Discussion: Our BLAST analysis revealed that our bacteria fell under the Serratia genus. The most common species of Serratia is marcescens: a gramnegative, facultatively-anaerobic bacterium and opportunistic pathogen that causes those pink rings around bath drains thanks to the red pigment prodigiosin (Haddix, 2018., Nazzaro, 2019).

The closest BLAST result was Serratia inhibens, a strain with anti-fungal properties that was discovered in 2020 (Hennessy, et al., 2020).

The Indole test indicated that our bacteria was unable to metabolize tryptophan [figure 3]. The urease reaction showed that the bacteria can break down urea [figure 6]. The catalase test indicated that our unknown can break down hydrogen peroxide [figure 11].

The TSI test shows that the bacteria creates hydrogen gas and ferments glucose [figure 7]. The oxidase test was negative meaning the unknown bacteria had no enzyme to break down the reagent [figure 12]. It can use citrate as a carbon source as revealed by the citrate test and uses flagella for movement as seen with the motility agar [figure 5, 4].

The MAC and EMB plates allowed us to determine that our bacteria is a lactose fermenter, which is rare for the genus (Enterobacteriaceae, 2020) [figures 8, 9].

Summary of Contradictions

The TSI test [figure 7] says it can't ferment lactose or sucrose. This contradicts the MAC and EMB while being supported by the XLD plate. Due to the strong results on the MAC and EMB, we believe the unknown Serratia strain can ferment lactose but further testing is required to form a solid conclusion.

MAC and EMB show lactose fermentation, but the XLD plate shows no sugar fermentation – this contradicts the MAC, EMB, and the glucose fermentation claims of the TSI.

Conclusion: We found two antibiotic producers and a third non-antibiotic producer that shows us the importance of performing several different tests in order to possibly discover unique strain-specific capabilities.

M Bennett, J., & Greene, M. (2020). Enterobacteriaceae. Science Direct. https://www.sciencedirect.com/topics/immunology-and-microbiology/ serratia-marcescens#:~:text=Serratia%20Species&text=Serratia%20strains%20are%20motile%2C%20rarely,appear%20to%20be%20acquired%20 exogenously. Haddix, P. L., & Shanks, R. M. Q. (2018). Prodigiosin pigment of Serratia marcescens is associated with increased biomass production. Archives of microbiology, 200(7), 989–999. https://doi.org/10.1007/s00203-018-1508-0 Hennessy, R. C., Dichmann, S. I., Martens, H. J., Zervas, A., & Stougaard, P. (2020). Serratia inhibens sp. nov., a new antifungal species isolated from potato (Solanum tuberosum). International journal of systematic and evolutionary microbiology, 70(7), 4204–4211. https://doi.org/10.1099/ijsem.0.004270 Nazzaro G. (2019). Etymologia: Serratia marcescens. Emerging Infectious Diseases, 25(11), 2012. https://doi.org/10.3201/eid2511.ET2511 Paper texture: https://unsplash.com/photos/k4Lt0CjUnb0